

This article was downloaded by: [Renmin University of China]

On: 13 October 2013, At: 10:34

Publisher: Taylor & Francis

Informa Ltd Registered in England and Wales Registered Number: 1072954 Registered office: Mortimer House, 37-41 Mortimer Street, London W1T 3JH, UK



Journal of Coordination Chemistry

Publication details, including instructions for authors and subscription information:

<http://www.tandfonline.com/loi/gcoo20>

Heterocyclic substituted thiosemicarbazides and their Cu(II) complexes: synthesis, spectral characterization, thermal, molecular modeling, and DNA degradation studies

O.A. El-Gammal^a, G.M. Abu El-Reash^a, S.E. Ghazy^a & T. Yousef^b

^a Department of Chemistry, Faculty of Science, Mansoura University, Mansoura 35516, Egypt

^b Department of Toxic and Narcotic Drug, Forensic Medicine, Mansoura Laboratory, Medicolegal Organization, Ministry of Justice, Egypt

Published online: 05 Apr 2012.

To cite this article: O.A. El-Gammal, G.M. Abu El-Reash, S.E. Ghazy & T. Yousef (2012) Heterocyclic substituted thiosemicarbazides and their Cu(II) complexes: synthesis, spectral characterization, thermal, molecular modeling, and DNA degradation studies, Journal of Coordination Chemistry, 65:10, 1655-1671, DOI: [10.1080/00958972.2012.674519](https://doi.org/10.1080/00958972.2012.674519)

To link to this article: <http://dx.doi.org/10.1080/00958972.2012.674519>

PLEASE SCROLL DOWN FOR ARTICLE

Taylor & Francis makes every effort to ensure the accuracy of all the information (the "Content") contained in the publications on our platform. However, Taylor & Francis, our agents, and our licensors make no representations or warranties whatsoever as to the accuracy, completeness, or suitability for any purpose of the Content. Any opinions and views expressed in this publication are the opinions and views of the authors, and are not the views of or endorsed by Taylor & Francis. The accuracy of the Content should not be relied upon and should be independently verified with primary sources of information. Taylor and Francis shall not be liable for any losses, actions, claims, proceedings, demands, costs, expenses, damages, and other liabilities whatsoever or howsoever caused arising directly or indirectly in connection with, in relation to or arising out of the use of the Content.

This article may be used for research, teaching, and private study purposes. Any substantial or systematic reproduction, redistribution, reselling, loan, sub-licensing, systematic supply, or distribution in any form to anyone is expressly forbidden. Terms & Conditions of access and use can be found at <http://www.tandfonline.com/page/terms-and-conditions>

Heterocyclic substituted thiosemicarbazides and their Cu(II) complexes: synthesis, spectral characterization, thermal, molecular modeling, and DNA degradation studies

O.A. EL-GAMMAL*[†], G.M. ABU EL-REASH[†], S.E. GHAZY[†] and T. YOUSEF[‡]

[†]Department of Chemistry, Faculty of Science, Mansoura University,
Mansoura 35516, Egypt

[‡]Department of Toxic and Narcotic Drug, Forensic Medicine, Mansoura Laboratory,
Medicolegal Organization, Ministry of Justice, Egypt

(Received 10 August 2011; in final form 17 February 2012)

Four Cu(II) complexes of *N*¹-phenyl-*N*²-(pyridin-2-yl)hydrazine-1,2-bis(carbothioamide) (H₂PPS), *N*-phenyl-2-(2-(pyridin-2-ylcarbamoithiyl)hydrazinyl)-2-thioxoacetamide (H₂PBO), *N*-phenyl-2-(pyridin-2-ylcarbamoithiyl)hydrazinecarboxamide (H₂APO), and 1-(amino-*N*-(pyridin-2-yl)methanethio)-4-(pyridin-yl)thiosemicarbazide (H₂PPY) have been prepared and characterized by elemental analyses, spectral (infrared (IR), UV-Visible, ¹H NMR, and electron spin resonance (ESR)) as well as magnetic and thermal measurements. Varying the substituents on the thiosemicarbazide led to remarkable modifications of the mode of coordination. IR spectral data reveal that the ligands are SN bidentate, NON tridentate, or NSNS tetradentate chelates forming structures in which copper is square-planar or octahedral. ESR spectra of these complexes are quite similar and exhibit an axially symmetric *g*-tensor parameter with $g_{\parallel} > g_{\perp} > 2.0023$ revealing an appreciable covalency with $d_{(x^2-y^2)}$ as the ground-state. Bond lengths, bond angles, HOMO, LUMO, and dipole moments have been calculated to confirm the geometry of the thiosemicarbazide derivatives and their corresponding complexes. Proton-ligand dissociation and copper-ligand stability constants of all ligands were calculated pH-metrically. The effect of the compounds on calf thymus DNA was investigated.

Keywords: Heterocyclic thiosemicarbazides; Copper(II) complexes; Thermal degradation; Calf thymus DNA

1. Introduction

Thiosemicarbazones are sulfur ligating systems with a range of biological properties [1–4]. Thiosemicarbazide syntheses can be modified from the parent aldehyde or ketone, or by substitution at ²C or ¹N [5–7]. Thiosemicarbazide complexes have anticancer and antimicrobial activity owing to their ability to diffuse through the semipermeable membrane of cell lines [5, 8–10]. The biological activity of copper, a metal that plays an important role in many biological processes [11], has been the subject of numerous studies [12–14]. Since Sigman *et al.* [15] discovered in 1979 that

*Corresponding author. Email: olaelgammal@yahoo.com

copper ions complexed to 1,10-phenanthroline were capable of cleaving DNA [16], copper complexes with a nitrogen donor heterocyclic ligand have been used widely to improve nuclease activity [17–21], mostly because of their ability to break the DNA chain in the presence of H_2O_2 and reducing agents [22]. In this work, we study the coordination chemistry of substituted thiosemicarbazides, N^1 -phenyl- N^2 -(pyridin-2-yl)hydrazine-1,2-bis(carbothioamide) (H_2PPS), N -phenyl-2-(pyridin-2-ylcarbamothioyl) hydrazinecarboxamide (H_2PBO), 1-(amino(thioformyl)- N -phenylform)-4-(pyridin-2-yl)thiosemicarbazide (H_2APO), and 1-(amino- N -(pyridin-3-yl)methanethio)-4-(pyridin-yl)thiosemicarbazide (H_2PPY) to correlate their structures with their ability to degrade DNA.

2. Experimental

2.1. Materials and physical measurements

All chemicals were purchased from Aldrich and Fluka and used without purification. Elemental analyses (C, H, and N) were performed on a Perkin-Elmer 2400 analyzer. All compounds were within $\pm 0.5\%$ of the theoretical values. Melting points were determined in open capillaries on an electrothermal melting point apparatus (Electrothermal Engineering Ltd., Essex, UK) and are uncorrected. ^1H NMR spectra were recorded in DMSO-d_6 on a JEOL JNM LA 300 WB spectrometer at 300 MHz using TMS (tetramethylsilane) as an internal standard (chemical shift in δ ppm) and the infrared (IR) spectra of the ligands were recorded as KBr discs on a Mattson 5000 FTIR spectrophotometer. Magnetic susceptibilities were measured with a Sherwood scientific magnetic susceptibility balance at 298 K. Thermogravimetric measurements (TGA, DTA, 20–1000°C) were recorded on a DTG-50 Shimadzu thermogravimetric analyzer at a heating rate of $10^\circ\text{C min}^{-1}$ and nitrogen flow rate of 20 mL min^{-1} . Electron spin resonance (ESR) spectra were obtained on a Bruker EMX spectrometer working in the X-band (9.78 GHz) mode with 100 kHz modulation frequency. The microwave power and modulation amplitudes were set at 1 mW and 4 Gauss, respectively. The low field signal was obtained after four scans with 10-fold increase in the receiver gain. A powder spectrum was obtained in a 2 mm quartz capillary at room temperature.

2.2. Synthesis of the ligands

4-(2-Pyridyl)-3-thiosemicarbazide (**1**) was synthesized as reported earlier [23]. Derivatives (**2–5**) were prepared by heating ethanolic solution of 4-(2-pyridyl)-3-thiosemicarbazide (**1**) (1.6 g, 100 mmol) with an equimolar amount of phenyl isothiocyanate, phenyl isocyanate, benzoyl isothiocyanate, and 2-pyridyl isothiocyanate, respectively.

2.2.1. H_2PPS (2**).** Yield 78%; m.p.: 178–180°C, $\text{C}_{13}\text{H}_{13}\text{N}_5\text{S}_2$ (303.41): Calcd C 51.46, H 4.32, N 23.08, S 21.14; found C 51.13, H 4.25, N 22.96, S 21.11. IR (KBr): $\nu/\delta = 1223$,

850 (C=S), 3099 (NH)_{a,a'}, 3160 (NH)_{b,c}, 1569 (C=N)_{py}, and 1598 (C=C). ¹H NMR δ = 15.4 (s, 1H, SH), 11.10, 10.97 (s, 1H, NH_{a,a'}), 8.27, 8.25 (d, J = 1.2 Hz, 1H, NH_{b,c}), 8.26 (d, J = 5.4 Hz, 1H, pyridine H₆), 7.85 (dd, J = 6.9, 5.4 Hz, 1H, pyridine H₄), 7.65 (dd, J = 6.9, 4.8 Hz, 1H, pyridine H₅), 7.37–7.50 (d, J = 5.4 Hz, 1H, pyridine H₃), 7.00–7.35 (m, 5H, Ph ring).

2.2.2. H₂PBO (3). Yield 79%; m.p.: 267–270°C, C₁₄H₁₃N₅OS₂ (331.42): Calcd C 50.74, H 3.95, N 21.13, O 4.83, S 19.35; found C 50.35, H 3.47, N 21.78, O 4.71, S 19.31. IR (KBr): ν/δ = 1243, 865 (C=S), 3158, 3100 (NH)_{a,a'}, 3239 (NH)_{b,c}, 1542 (C=N)_{py}, 1675 (C=O), and 1604 (C=C). ¹H NMR δ = 11.09, 10.53 (s, 1H, NH_{a,a'}), 9.54, 8.20 (d, 1H, NH_{b,c}), 8.27 (d, J = 8.1 Hz, 1H, pyridine H₃), 2 (d, J = 5.1 Hz, 1H, pyridine H₆), 7.55 (dd, J = 6.3, 5.7 Hz, 1H, pyridine H₄), 7.78 (dd, J = 7.5, 6.3 Hz, 1H, pyridine H₅), 7.12–7.23 (m, 5H, Ph ring).

2.2.3. H₂APO (4). Yield 81%; m.p.: 180–182°C, C₁₃H₁₃N₅OS (287.34): Calcd C 54.34, H 4.56, N 24.37, O 5.57, S 11.16; found C 54.15, H 4.45, N 23.98, O 5.78, S 11.11. IR (KBr): ν/δ = 1247, 800 (C=S), 3170, 3104 (NH)_{a,a'}, 3220 (NH)_{b,c}, 1641 (C=N)_{py}, 1672 (C=O), and 1600 (C=C). ¹H NMR: δ = 15.8 (s, 1H, SH), δ = 11.9, 11.30 (s, 1H, NH_{a,a'}), 8.36, 8.09 (d, 1H, NH_{b,c}), 8.23 (dd, J = 8.7, 6 Hz, 1H, pyridine H₅), 8.01 (d, J = 5.3 Hz, 1H, pyridine H₆), 7.9 (d, J = 6.4 Hz, 1H, pyridine H₃), 7.66 (dd, J = 6.1, 4.8 Hz, 1H, pyridine H₄), 7.0–7.64 (m, 5H, Ph ring).

2.2.4. H₂PPY (5). Yield 82%; m.p.: 307–310°C, C₁₂H₁₂N₆S₂ (304.06): Calcd C 47.35, H 3.97, N 27.61, S 21.07; found C 47.02, H 4.05, N 27.86, S 21.31. IR (KBr): ν/δ = 1218, 875 (C=S), 3174 (NH)_{a,a'}, 3234 (NH)_{b,c}, 1562 (C=N)_{py}, and 1604 (C=C). ¹H NMR δ = 15.4 (s, 1H, SH), 11.10 (s, 1H, NH_{a,a'}), 8.29, 8.28 (d, J = 3.9, 1H, NH_{b,c}), 8.21 (d, J = 6.2 Hz, 1H, pyridine H₆), 8.17 (d, J = 7.4 Hz, 1H, pyridine H₃), 7.88 (dd, J = 5.5, 3.3 Hz, 1H, pyridine H₅), 7.85 (dd, J = 6.6, 5.4 Hz, 1H, pyridine H₄).

2.3. Synthesis of Cu(II) complexes

Ethanol solution of thiosemicarbazides (1.0 mmol) and hydrated copper chloride (1.0 mmol) was refluxed for 2–3 h. The precipitates that formed were filtered off, washed with hot ethanol followed by diethyl ether, and dried in a vacuum desiccator over anhydrous CaCl₂. The complexes have high melting points and are insoluble in common organic solvents and soluble in DMF or DMSO.

2.3.1. [Cu(HPPS)Cl(H₂O)₂] (6). Anal. Calcd for C₁₃H₁₄CuN₅OS₂Cl: C 37.23, H 3.36, N 16.69, S 15.29, Cu 15.2, Cl 8.45; Found: C 37.59, H 3.18, N 17.05, S 15.43, Cu 15.4, Cl 8.15. Main IR peaks (KBr, cm⁻¹): ν (NH)_{a,a'} 3120, 3070; ν (NH)_{b,c} 3230; ν (C=N)_{py} 1616; ν/δ (C=S)_{a,a'} 1232, 860; ν (C=N)* 1600; ν (C-S) 646; UV-Vis (DMF, cm⁻¹) 35,211.

2.3.2. [Cu(HPBO)Cl(H₂O)₂] (7). Anal. Calcd for C₁₄H₁₆CuN₅O₃S₂Cl: C 36.12, H 3.46, N 15.04, S 13.77, Cu 13.6, Cl 4.61; Found: C 36.33, H 3.78, N 15.55, S 13.48, Cu 13.4, Cl 7.81. Main IR peaks (KBr, cm⁻¹): ν (NH)_{a,a'} 3187; ν (C=O) 1673; ν (C=N)_{py}

1562; ν/δ (C=S)_{a,a'} 1221, 854; ν (C=N)* 1612; ν (C-S) 646; UV-Vis (DMF, cm^{-1}) 32,258; 28,409; 18,050.

2.3.3. [Cu(HAPO)Cl](H₂O) (8). Anal. Calcd for C₁₃H₁₈CuN₅O₄SCl: C 35.54, H 4.13, N 15.04, S 13.77, Cu 14.50, Cl 8.10; Found: C 35.66, H 4.05, N 15.55, S 13.48, Cu 14.45, Cl 8.30. Main IR peaks (KBr, cm^{-1}): ν (NH)_{a,a'} 3244; ν (NH)_{b,c} 3115; ν (C=N)_{py} 1621; ν/δ (C=S)_{a,a'} 1232, 865; ν (C=N)* 1567; UV-Vis (DMF, cm^{-1}) 35,211; 32,467; 27,624; 23,585; 19,685; 15,291.

2.3.4. [Cu(HPPY)Cl (H₂O)₂] (9). Anal. Calcd for C₁₂H₁₅CuN₆O₂S₂Cl: C 32.84, H 3.45, N 19.16, S 14.63, Cu 14.4, Cl 8.1; Found: C 32.63, H 3.8, N 19.34, S 14.28, Cu 13.4, Cl 8.8. Main IR peaks (KBr, cm^{-1}): ν (NH)_{a,a'} 3187; ν (C=N)_{py} 1562; ν/δ (C=S)_{a,a'} 1221, 840; ν (C=N)* 1612; ν (C-S) 656; UV-Vis (DMF, cm^{-1}) 35,211; 31,847; 29,585; 25,126; 19,157.

2.4. pH metric studies

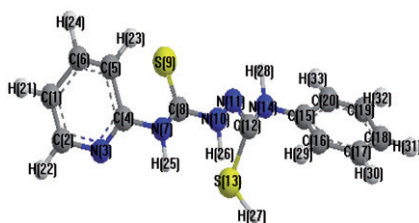
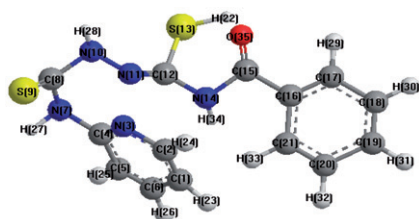
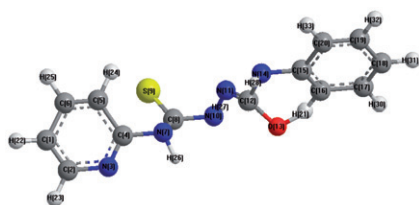
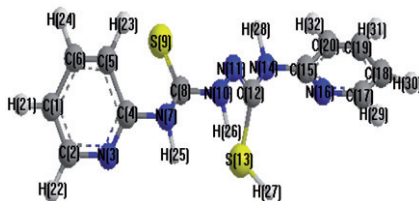
The experimental procedure involves pH-metric titrations of solution mixtures against standardized NaOH ($9.2 \times 10^{-3} \text{ mol L}^{-1}$) in 50% (v/v) dioxane-water at (298, 303, and 308 K) and at constant ionic strength of ($\mu = 0.1, 0.15, \text{ and } 0.2 \text{ mol L}^{-1} \text{ KCl}$). The solution mixtures (i–iii) were prepared as follows:

- (i) 1.25 mL ($1.1 \times 10^{-2} \text{ mol L}^{-1}$) HCl + 1.25 mL (0.1, 0.15, and 0.2 mol L^{-1}) KCl + 10 mL twice-distilled H₂O.
- (ii) 1.25 mL ($1.1 \times 10^{-2} \text{ mol L}^{-1}$) HCl + 1.25 mL (0.1, 0.15, and 0.2 mol L^{-1}) KCl + 2.5 mL ($5 \times 10^{-3} \text{ mol L}^{-1}$) H₂PPS, H₂PBO, H₂APO, and H₂PPY + 10 mL twice-distilled H₂O.
- (iii) 1.25 mL ($1.1 \times 10^{-2} \text{ mol L}^{-1}$) HCl + 1.25 mL (0.1, 0.15, and 0.2 mol L^{-1}) KCl + 2.5 mL ($5 \times 10^{-3} \text{ Cu}^{2+}$ solution) + 0.5 mL ($5 \times 10^{-3} \text{ mol L}^{-1}$) metal ion in twice-distilled water + 9.5 mL H₂O.

The total volume was adjusted to 25 mL by adding dioxane in each case. The pH-meter readings in 50% (v/v) dioxane mixture are corrected according to the Van Uitert and Hass relation [24].

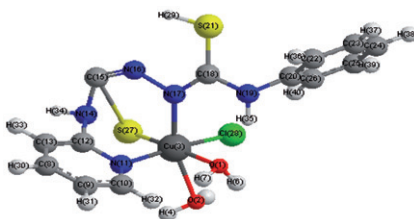
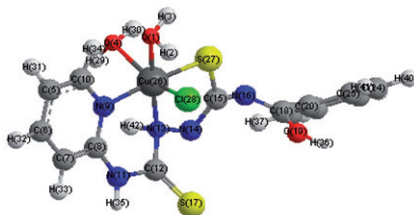
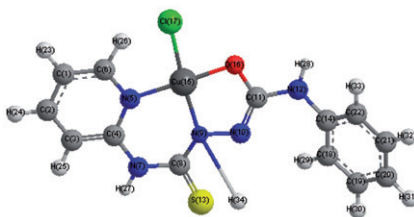
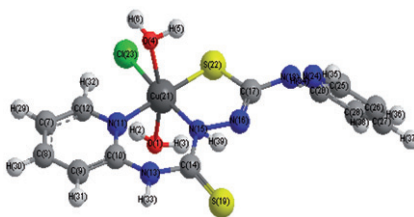
2.5. DNA interaction studies

A solution of 2 mg of calf thymus DNA (ct-DNA) was dissolved in 1 mL of sterile distilled water. Stock concentrations of the ligands and complexes were prepared by dissolving 2 mg mL^{-1} in DMSO. Equal volumes from each compound and DNA were mixed thoroughly and kept at room temperature for 2–3 h. The effect of the chemicals on the DNA was analyzed by agarose gel electrophoresis. Loading dye ($2 \mu\text{L}$) was added to $15 \mu\text{L}$ of the DNA mixture before being loaded into the well of agarose gel. The loaded mixtures were fractionated by electrophoresis, visualized by UV and photographed [25].

Scheme 1. Molecular modeling of H₂PPS.Scheme 2. Molecular modeling of H₂PBO.Scheme 3. Molecular modeling of H₂APO.Scheme 4. Molecular modeling of H₂PPY.

2.6. Molecular modeling

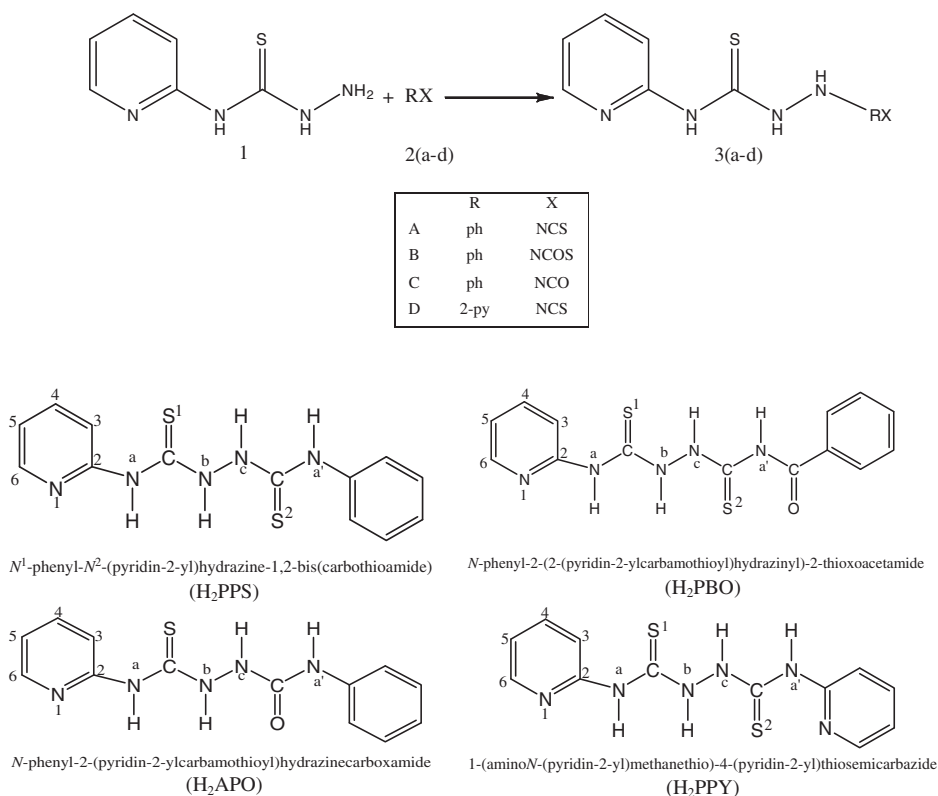
To gain better insight on the molecular structure of the ligands (schemes 1–4) and complexes (schemes 5–8), geometric optimization and conformational analysis have been performed using MM+ [26] force field as implemented in HyperChem 8 [27].

Scheme 5. Molecular modeling of $[\text{Cu}(\text{HPPS})\text{Cl}(\text{H}_2\text{O})_2]$ complex.Scheme 6. Molecular modeling of $[\text{Cu}(\text{HPBO})\text{Cl}(\text{H}_2\text{O})_2]$ complex.Scheme 7. Molecular modeling of $[\text{Cu}(\text{HAPO})\text{Cl}](\text{H}_2\text{O})$ complex.Scheme 8. Molecular modeling of $[\text{Cu}(\text{HPPY})\text{Cl}(\text{H}_2\text{O})_2]$ complex.

3. Results and discussion

3.1. *Synthesis and spectroscopy*

In continuation of our previous work on pyridine chemistry [28], we synthesized some new thiosemicarbazide derivatives containing the pyridine ring (scheme 9).



3.2. IR spectra

IR spectra of the free ligands show three bands in the region $3234\text{--}3100\text{ cm}^{-1}$ attributable to stretching modes of the $(\text{NH})_{\text{a,a}'}$ and $\nu(\text{NH})_{\text{b,c}}$ groups. The stretching and out-of-plane ring deformation modes (ν/δ) of the pyridine ring are observed at ≈ 1560 and $620\text{--}635\text{ cm}^{-1}$ [29]. Strong bands due to stretching vibration of $\nu(\text{C}=\text{N})$ (azomethine) in H_2PPS and H_2PBO thiosemicarbazides are at 1646 and 1641 cm^{-1} , respectively; this band was difficult to recognize in H_2PPY because it is overlapped with $\nu(\text{C}=\text{C})$ of the pyridine ring. The stretching and bending vibrations (ν/δ) of the $\text{C}=\text{S}$ groups are observed as strong bands at $1218\text{--}1247\text{ cm}^{-1}$ and $840\text{--}865\text{ cm}^{-1}$, respectively [30, 31]. As these ligands contain two $\text{C}=\text{S}$ groups, $(\text{C}=\text{S})^1$ and $(\text{C}=\text{S})^2$, we propose that $(\text{C}=\text{S})^1$ is in the thione form and $(\text{C}=\text{S})^2$ in the thiol form as confirmed by the existence of additional bands at ~ 2600 and 660 cm^{-1} in the spectra of the aforementioned ligands assignable to $\nu(\text{C}\text{--}\text{S})$ and $\nu(\text{SH})$. This assumption is further supported by the absence of bands due to $\nu(\text{C}=\text{N})$ (azomethine), $\nu(\text{C}\text{--}\text{S})$, and $\nu(\text{SH})$ in the IR spectrum of the starting (4-pyridyl thiosemicarbazide) [32] and the absence of the SH signal in the ^1H NMR spectrum of H_2APO .

Also, the appearance of $\delta(\text{C}\text{--}\text{S})$ in H_2PPY at higher wavenumber, 683 cm^{-1} , than that of other ligands may be due to presence of pyridyl groups which act as electron-withdrawing groups.

In H₂APO, the sharp band at 3521 cm⁻¹ may be due to $\nu(\text{OH})$ in addition to the $\nu(\text{CO})$ band at 1675 cm⁻¹, suggesting the keto–enol tautomerism.

IR spectra of the investigated Cu(II) complexes show that H₂PPS, H₂PBO, H₂APO, and H₂PPY coordinate as mono-negative tridentate through (C=N) of pyridine, N of NH_b, and one of the (C=S) groups adjacent to N⁴ in the thiol form (H₂PPS, H₂PBO, and H₂PPY) or enolized carbonyl group in H₂APO.

This chelation is supported by the shift of deformation mode of the pyridine ring either to lower or higher wavenumber, indicative of coordination of the metal to pyridine [33], and the band due to thioamide(4) at 840–875 either shifts to lower wavenumber [34] or disappears with simultaneous appearance of a new band assignable to $\nu(\text{C–S})$, suggesting coordination *via* C=S either in the thione form [Cu(HPPS)Cl(H₂O)₂] or the thiol form [Cu(HPBO)Cl(H₂O)₂] and [Cu(HPPY)Cl(H₂O)₂] [35]. The shift of $\nu(\text{N–N})$ to higher wavenumber is further evidence for coordination *via* (NH)_b. In the IR spectrum of [Cu(HAPO)Cl](H₂O), CS bands at the same position suggest that they do not participate in coordination and the disappearance of the band at 3363 cm⁻¹ due to the (C–OH) mode, in addition to a new band at 533 cm⁻¹ due to $\nu(\text{Cu–O})$, reveals this mode of coordination.

In all the complexes, a band at 480–424 cm⁻¹ assigned to $\nu(\text{Cu–N})$ supports the proposed mode of coordination. IR spectra of all complexes exhibit ν_{asym} and $\nu_{\text{sym}}(\text{OH})$ and $\Delta(\text{H}_2\text{O})$ vibrations at $\approx 3399\text{--}3450\text{ cm}^{-1}$ and a weak shoulder at 1625 cm⁻¹, respectively, suggesting water [36]. Broad bands at 875–850 cm⁻¹ and $\approx 540\text{--}550$ refer to P_f(H₂O) and P_w(H₂O) vibrations and support that water is coordinated, confirmed by TGA studies.

3.3. ¹H NMR spectra

¹H NMR spectra of H₂PPS, H₂PBO, H₂APO, and H₂PPY in DMSO-d₆ show signals at $\delta = (10.30\text{--}10.90)$, $(11.09\text{--}11.90)$, $(8.09\text{--}8.25)$, $(8.27\text{--}9.54)$, and $(15.40, 15.80)$ ppm assignable to (NH)_{a,a'}, (NH)_{b,c}, and SH protons, respectively. The peak of (NH)_{a,a'} disappears and the intensity of the (NH)_{b,c} peak decreases on addition of D₂O, which suggests that they are easily exchangeable. These protons shift downfield because they hydrogen bond with N of pyridine and S of CS. The protons of (NH)_a and (NH)_{a'} appear as singlets as expected since the NH protons are decoupled from nitrogen and protons from adjacent atoms. (NH)_b and (NH)_c show coupling with the adjacent hydrogen as doublets. This coupling can be attributed to the low NH exchange rate. Signals attributed to pyridyl appear at $\delta = 8.26$ (d, $J = 5.40$ Hz, 1H, pyridine H6), 7.85 (dd, $J = 6.9, 5.4$ Hz, 1H, pyridine H4), 7.65 (dd, $J = 6.9, 4.8$ Hz, 1H, pyridine H5), and 7.35–7.50 (d, $J = 5.4$ Hz, 1H, pyridine H3) for H₂PPS, 8.23 (dd, $J = 8.7$ Hz, 1H, pyridine H5), 8.01 (d, $J = 5.3$ Hz, 1H, pyridine H6), 7.9 (d, $J = 6.4$ Hz, 1H, pyridine H3), 7.66 (dd, $J = 6.1$ Hz, 1H, pyridine H4) for H₂PBO, 8.23 (dd, $J = 8.7$ Hz, 1H, pyridine H5), 8.01 (d, $J = 5.3$ Hz, 1H, pyridine H6), 7.9 (d, $J = 6.4$ Hz, 1H, pyridine H3), 7.66 (dd, $J = 6.1$ Hz, 1H, pyridine H4) for H₂APO, and 8.21 (d, $J = 6.2$ Hz, 1H, pyridine H6), 8.17 (d, $J = 7.4$ Hz, 1H, pyridine H3), 7.88 (dd, $J = 5.5$ Hz, 1H, pyridine H5), 7.85 (dd, $J = 7.3$ Hz, 1H, pyridine H4) for H₂PPY [37]. The signal at $\delta(8.01\text{--}8.22)$ ppm for pyridine H6 in all thiosemicarbazides is superimposed with that of some NH (NH)_{b,c} protons. Also, spectra of H₂PPS, H₂PBO, and H₂APO at 7.00–7.64 ppm due to phenyl protons appear as complicated multiplets. A signal at $\delta 15.4$ ppm in the spectrum of

Table 1. Magnetic moment and electronic spectra of the Cu(II) complexes of H₂PPS, H₂PBO, H₂APO, and H₂PPY.

Compound	Solvent	Band position (cm ⁻¹)	μ_{eff} (B.M.)
H ₂ PPS	DMF	35,211; 31,847; 29,069	–
	Nujol	–	
[Cu(HPPS)Cl(H ₂ O) ₂]	DMF	35,211; 28,089; 26,178 18,587; 16,077	1.78
	Nujol	23,364; 20,408; 17,301	
H ₂ PBO	DMF	35,461; 32,680; 29,070	–
	Nujol	–	
[Cu(HPBO)Cl(H ₂ O) ₂]	DMF	32,258; 28,409; 18,050	2.03
	Nujol	25,000; 30,675; 17,606	
H ₂ APO	DMF	35,211; 32,051	–
	Nujol	–	
[Cu(HAPO)Cl](H ₂ O)	DMF	35,211; 32,467; 27,624	1.54
	Nujol	23,585; 19,685; 15,291	
H ₂ PPY	DMF	35,971; 28,409; 22,321	–
	Nujol	35,211; 30,120; 18,050	
[Cu(HPPY)Cl(H ₂ O) ₂]	Nujol	35,211; 31,847; 29,585	1.83
	DMF	25,126; 19,157	

H₂PPY due to SH and a signal at δ 12.80 ppm due to OH in H₂APO confirm the thiol-enol tautomerism in solution.

3.4. Electronic spectra and magnetic moments

The absorption bands and the magnetic moments of the thiosemicarbazides and their Cu(II) complexes in DMF and Nujol mull are recorded in table 1. The spectra exhibit bands at 35,111–35,971 cm⁻¹ and 30,120–32,680 cm⁻¹ assignable to $\pi \rightarrow \pi^*$ and $n \rightarrow \pi^*$ transitions, respectively [38]. Charge transfer is observed in spectra of complexes with a new $n \rightarrow \pi^*$ band at 28,089–29,585 cm⁻¹. The charge transfer band assigned to (S \rightarrow Cu) [38, 39] in [Cu(HPPS)Cl(H₂O)₂], [Cu(HPBO)Cl(H₂O)₂], and [Cu(HPPY)Cl(H₂O)₂] appeared at higher energies (22,320–25,000 cm⁻¹) in Nujol possibly due to coordination *via* sulfur. The d–d transition bands for [Cu(HPPS)Cl(H₂O)₂], [Cu(HPBO)Cl(H₂O)₂], and [Cu(HPPY)Cl(H₂O)₂] at 18,050–19,685 cm⁻¹ in DMF and at 17,301–18,116 cm⁻¹ in Nujol are consistent with octahedral complexes. For [Cu(HAPO)Cl](H₂O), the band due to d–d transition appeared at 15291 cm⁻¹ in DMF confirming a square-planar environment. These geometries will be further confirmed by ESR spectra. Magnetic moment values fall in the range reported for d⁹ Cu(II) complexes.

3.5. ESR spectra

ESR spectra of Cu(II) complexes provide information about hyperfine and super-hyperfine structures which are important in studying the metal ion environment in

Table 2. ESR data of some Cu(II) complexes at room temperature.

Complex	g_{\parallel}	g_{\perp}	$A_{\parallel} \times 10^{-4} \text{ (cm}^{-1}\text{)}$	G	$g_{\parallel}/A_{\parallel}$	α^2	β^2
[Cu(HPPS)Cl(H ₂ O)]	2.28	2.06	165	4.60	138	0.80	0.90
[Cu(HPBO)Cl(H ₂ O)](H ₂ O)	2.29	2.06	168	3.30	136	0.73	0.74
[Cu(HAPO)Cl](H ₂ O) ₃	2.25	2.08	–	3.12	–	–	–
[Cu(HPPY)Cl(H ₂ O)](H ₂ O)	2.23	2.07	150	3.20	148	0.71	0.87

complexes, i.e., the geometry, nature of the ligation sites and degree of covalency of the metal ligand bonds (Supplementary material). The spin Hamiltonian parameters of the complexes with Cu(II), $S = 1/2$, $I = 3/2$, were calculated and are summarized in table 2.

Room temperature solid state ESR spectra of these complexes are similar and exhibit axially symmetric g -tensor parameters with $g_{\parallel} > g_{\perp} > 2.0023$ revealing an appreciable covalency with $d_{(x^2-y^2)}$ as the ground-state [40], characteristic of square-planar, square-pyramidal or octahedral stereochemistry [41].

In axial symmetry the g -values are related by the expression $G = (g_{\parallel} - 2)/(g_{\perp} - 2) = 4$, where G is the exchange interaction parameter. According to Hathaway and Billing [42], if G is greater than 4, the exchange interaction between copper(II) centers in the solid state is negligible, whereas when it is less than 4, considerable exchange interaction is indicated. The calculated G values (Supplementary material) show $G > 4$ for [Cu(HPPS)Cl(H₂O)]₂ and less than 4 for [Cu(HPBO)Cl(H₂O)]₂, [Cu(HAPO)Cl](H₂O), and [Cu(HPPY)Cl(H₂O)]₂ [43].

A band corresponding to forbidden magnetic dipolar transition for [Cu(HPBO)Cl(H₂O)]₂, [Cu(HPPS)Cl(H₂O)]₂, and [Cu(HPPY)Cl(H₂O)]₂ is not observed at half-field (*ca* 1600 G, $g = 4.0$), revealing mononuclear Cu(II) complexes [44]. Superhyperfine structures for these complexes are not seen at higher fields excluding any interaction of the nuclear spins of nitrogen ($I = 1$) with the unpaired electron density on Cu(II).

The absorption for [Cu(HAPO)Cl](H₂O) consists of a broad band centered at $g = 2$, attributable to dipolar broadening and enhanced spin lattice relaxation [45]. This line broadening is probably due to insufficient spin exchange narrowing toward the coalescence of four copper hyperfine lines to a single line; similar powder ESR line shapes have been observed for many square-planar or distorted octahedral dimeric Cu(II) complexes with strong intra-dimer spin-exchange interaction [46]. Hyperfine splitting due to the spin of copper was not detected because of the broadness. These observations confirm occurrence of super exchange coupling between Cu(II) ions in the dimeric complexes, consistent with the subnormal effective magnetic moment values which may refer to sulfur coordination [47].

The tendency of A_{\parallel} to decrease with concomitant increase of g_{\parallel} is an index of tetrahedral distortion in the coordination sphere of copper. To quantify the degree of distortion of the Cu(II) complexes, we selected $f(\alpha) = g_{\parallel}/A_{\parallel}$ obtained from the ESR spectra. Values of 110–120 are typical for planar complexes, while the range 130–150 is characteristic of slight to moderate distortion and 180–250 cm⁻¹ indicate considerable distortion [48, 49]. The ratio $g_{\parallel}/A_{\parallel}$ for [Cu(HPBO)Cl(H₂O)]₂, [Cu(HPPS)Cl(H₂O)]₂, and [Cu(HPPY)Cl(H₂O)]₂ are 136, 138, and 148, respectively, demonstrating significant dihedral angle distortion in the xy -plane, consistent with distorted octahedral geometry around the copper site.

Molecular orbital coefficients, α^2 (a measure of the covalency of the in-plane σ -bonding between a copper 3d orbital and the ligand orbitals) and β^2 (covalent in-plane π -bonding), were calculated by using the following equations [50–53]:

$$\alpha^2 = (A_{\parallel}/0.036) + (g_{\parallel} - 2.0023) + 3/7(g_{\perp} - 2.0023) + 0.04,$$

$$\beta^2 = (g_{\parallel} - 2.0023)E / -8\lambda\alpha^2,$$

where $\lambda = -828 \text{ cm}^{-1}$ for free copper ion and E is the electronic transition energy.

As a measure of the covalency of the in-plane σ -bonding, $\alpha^2 = 1$ indicates complete ionic character, whereas $\alpha^2 = 0.5$ denotes 100% covalent bonding with the assumption of negligibly small values of overlap integral. β^2 gives an indication of the covalency of the in-plane π -bonding. The smaller β^2 , the larger is the covalency of the bonding.

Values of α^2 and β^2 for $[\text{Cu}(\text{HPBO})\text{Cl}(\text{H}_2\text{O})_2]$, $[\text{Cu}(\text{HPPS})\text{Cl}(\text{H}_2\text{O})_2]$, and $[\text{Cu}(\text{HPPY})\text{Cl}(\text{H}_2\text{O})_2]$ indicate that the in-plane σ -bonding and in-plane π -bonding are covalent, which originates from thiolato sulfur binding incorporating greater covalency in the metal–ligand bonding through delocalized d_{π} – p_{π} in-plane π -bonding as evidenced from higher β^2 values [54]. Data for $[\text{Cu}(\text{HAPO})\text{Cl}](\text{H}_2\text{O})$ show that in-plane σ -bonding and in-plane π -bonding are ionic. For square-planar geometry, the lower values of β_2 compared to α_2 indicate that in-plane π -bonding is more covalent than the in-plane σ -bonding, consistent with other reported values [55–58].

3.6. Molecular modeling

The molecular structure and atom-numbering of ligands and its Cu(II) complex are shown in schemes 1–8.

From data recorded in Supplementary tables (S1–S9) for bond lengths and angles, one can conclude the following: (1) bond angles of the thiosemicarbazide are altered upon coordination with largest effects on S(9)–C(8)–N(10), C(8)–N(10)–N(11), N(10)–N(11)–C(12), and C(4)–N(7)–C(8) angles which are reduced from 123.3°, 116.8°, 123.3°, and 127.6° in H_2PPS to 97.6°, 127.5°, 107.5°, and 106° for $[\text{Cu}(\text{HPPS})\text{Cl}(\text{H}_2\text{O})_2]$; N(10)–N(11)–C(12) and N(11)–C(12)–S(13) from 124° and 126.1° in H_2PBO to 115° and 119.6° for $[\text{Cu}(\text{HPBO})\text{Cl}(\text{H}_2\text{O})_2]$; C(5)–C(4)–N(7) and N(10)–N(11)–C(12) from 126.6° and 122.5° in H_2APO to 121.6° and 114.3° for $[\text{Cu}(\text{HAPO})\text{Cl}](\text{H}_2\text{O})$, and N(11)–C(12)–S(13) and N(11)–C(12)–N(14) from 123.6° and 115.6° in H_2PPY to 118.4° and 122.5° for $[\text{Cu}(\text{HPPY})\text{Cl}(\text{H}_2\text{O})_2]$ [59]. (2) All bond angles in the complexes are close to octahedral geometry except for $[\text{Cu}(\text{HAPO})\text{Cl}](\text{H}_2\text{O})$, which has a square-planar geometry. (3) All groups taking part in coordination, C=S, N–H, (C=N)_{py}, and (C=O), have bonds longer than in the ligand. (4) The presence of two water molecules in *trans* position for $[\text{Cu}(\text{HPPY})\text{Cl}(\text{H}_2\text{O})_2]$. (5) Lower highest occupied molecular orbital (HOMO) energy values show weaker electron donation while higher HOMO energy implies a good electron donor. Lowest unoccupied molecular orbital energy represents the ability of a molecule receiving electron [60].

3.7. Thermogravimetric studies

The stages of decomposition, temperature range, decomposition product, and weight loss percentages of complexes are given in table 3. A glance at the table indicates that

Table 3. Decomposition steps with temperature range and weight loss for ligands and their copper complexes.

Compound	Temperature range (°C)	Removed species	Weight loss	
			Found (%)	Calcd (%)
H ₂ PPS	141–228	–Py + NH	30.84	30.68
	229–371	–Py + NH + CS	45.76	45.18
	513–691	–N ₂ H ₂	9.58	9.8
	691–800	CS (residue)	13.8	14.5
[Cu(HPPS)Cl(H ₂ O) ₂]	190–279	–2H ₂ O + HCl	16.78	16.57
	280–390	–Py	17.65	17.85
	391–527	–CN	5.85	5.95
	528–754	–CN ₂ H + NH + Ph	30.17	30.44
	754–800	CuS + S (residue)	29.55	29.19
H ₂ PBO	0–210	–Py + NH	28.58	28.09
	210–370	–C ₃ H ₃ S ₂ O	48.16	48.64
	370–700	C ₆ H ₅	23.28	23.27
[Cu(HPBO)Cl(H ₂ O) ₂]	188–292	–2H ₂ O	7.87	7.94
	292–441	–Cl + Py + N ₂ H ₂ + 2C	35.65	35.85
	441–695	–C ₆ H ₅ + NH + N + C	25.69	25.37
	695–800	CuO + 2S (residue)	30.79	30.84
H ₂ APO	126–217	–C ₅ H ₅ N ₂ + N ₂ H ₂	42.15	42.85
	217–344	–C ₆ H ₅ + CO + NH	41.11	41.82
	344–700	CS (residue)	16.74	15.34
[Cu(HAPO)Cl](H ₂ O)	21–137	–H ₂ O	4.03	4.47
	137–258	–Py + Cl + NH	31.77	31.88
	258–397	–C ₆ H ₅	19.63	19.12
	397–570	–NH + C	6.51	6.69
	570–776	–N ₂ H + C	10.93	10.17
	776–800	CuO + S (residue)	27.13	27.8
H ₂ PPY	188–246	–N ₂ H ₂	10.57	9.86
	246–400	–2Py + N ₂ H ₂	60.96	61.18
	400–700	2CS (residue)	28.78	28.96
[Cu(HPPY)Cl(H ₂ O) ₂]	290–467	–2H ₂ O + Cl	16.71	16.30
	467–660	–2Py + N ₄ H ₃ + 2C	54.16	54.56
	660–800	Cu + 2S (residue)	29.13	29.12

thermal stabilities of ligands increase on complex formation. TGA curves of [Cu(HPPS)Cl(H₂O)₂] as a representative example (Supplementary material) display five degradation steps. The first at 190–279°C with weight loss of 16.78 (Calcd 16.57%) is attributed to loss of the two lattice water molecules and Cl. The second step with weight loss of 17.65 (Calcd 17.85%) at 280–390°C corresponds to removal of pyridine. The third step at 391–557°C with weight loss of 5.85 (Calcd 5.95%) is for removal of CN. The fourth step (558–754°C) can be ascribed to elimination of CN₂H fragment with weight loss of 30.17 (Calcd 30.44%). The residual is CuS + S (found 29.55, Calcd 29.19%).

3.8. pH metric studies

The average number of protons associated with ligand \bar{n}_a was calculated at different pH-values using the Irving–Rossotti equation [61] and used to draw proton-ligand

formation curves, which indicated that the maximum n_a is ~ 2 revealing that the investigated ligands have two dissociable protons of NH^b and NH^c , respectively. An inspection of the results in table 4 reveals the following:

- (1) **Effect of substituent:** pK_1 values of the ligands are influenced by the inductive effect of the substituent. The pK values under the same conditions of temperature and ionic strength can be arranged as $\text{H}_2\text{APO} > \text{H}_2\text{PPS} > \text{H}_2\text{PPY} > \text{H}_2\text{PBO}$, attributed to hydrogen-bond formation. The first three compounds form two hydrogen bonds and the last one forms three hydrogen bonds, and as a result it is difficult to lose H^+ , so the pK is high. On substituting carbonyl by thione, the pK value decreases as oxygen is more electronegative than sulfur. The presence of pyridyl in H_2PPY (i.e., electron withdrawing) decreases the electron density more effectively than phenyl, giving a stronger hydrogen bond and, consequently, smaller pK .
- (2) **Effect of ionic strength:** There is an increase in pK with increasing ionic strength of the medium, consistent with that reported [62].
- (3) **Effect of temperature:** pK_a values decrease with increasing temperature, i.e., the acidity of ligands increase with increasing temperature.
- (4) **The stepwise stability constants of complexes:** The values (table 5) decrease with increasing temperature, indicating that complexation is unfavorable with increasing temperature.
- (5) **Thermodynamic parameters:** ΔG° values are positive, indicating non-spontaneous character of the dissociation processes. ΔS° values were negative, indicating that dissociation is entropically unfavorable due to disorder of the solvent around the ligand [63]. Positive values of ΔH° indicate that dissociation of ligands is endothermic.

“Supplementary material” shows that ΔG° values are negative, indicating spontaneous chelation. Negative values of ΔS° indicate that complexation is entropically unfavorable while negative values of ΔH° suggest that chelation is accompanied by generation of heat, indicating that the processes are exothermic, i.e., complexation is favorable at lower temperature.

3.9. DNA degradation studies

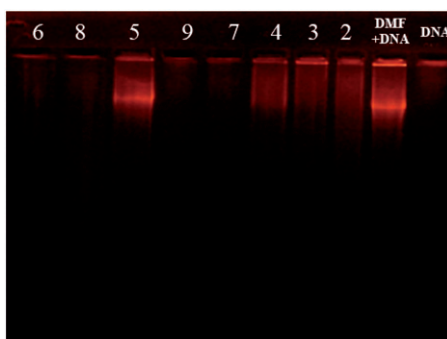
DNA cleavage reactions generally occur through three pathways, namely oxidative strand cleavage by abstraction of a sugar hydrogen (s), hydrolytic cleavage involving the phosphate, and by base oxidations primarily directed at guanine. Irradiation is one methodology for inducing DNA cleavage [64]. For the sulfur containing thiosemicarbazides thio or thione moieties exhibit efficient intersystem crossing to the triplet state $^3(n-\pi^*)$ and/or $^3(\pi-\pi^*)$. Such state with a longer lifetime can either cause damage to DNA directly or can activate oxygen from its triplet state ($^3\text{O}_2$) to the cytotoxic singlet state ($^1\text{O}_2$) [65, 66]. Examining DNA degradation assay for H_2PPS , H_2PBO , H_2APO , and H_2PPY , and their corresponding Cu(II) complexes revealed variability on their immediate damage on the ct-DNA (figure 1). The results show that either DNA in DMF (+ve control) or DNA treated with **2**, **3**, **4**, and **5** showed no effect through the incubation period, whereas **6** exhibited partial degradation, judged by the rate of DNA migration in the gel. DNA disappeared completely after treatment with **7**, **8**, and **9** due to direct contact of these complexes with DNA, which is necessary to degrade it. The

Table 4. The dissociation constants of H₂PPS in 50% (v/v) dioxane-water and KCl at different temperatures and their thermodynamic parameters.

Compound	μ (mol L ⁻¹)	Dissociation constant						Free energy change (ΔG) (kJ mol ⁻¹)						Enthalpy change (ΔH) (kJ mol ⁻¹)	Entropy change (ΔS) (J mol ⁻¹ K ⁻¹)		
		298 K		303 K		308 K		298 K		303 K		308 K					
		pK1	pK2	pK1	pK2	pK1	pK2	pK1	pK2	pK1	pK2	pK1	pK2				
H ₂ PPS	0.1	13.45	8.25	12.89	8.15	8.72	6.76	82.7	24.8	83.3	25.0	83.9	25.2	43.37	13.66	-131.9	-37.55
	0.15	13.44	10.2	11.98	8.56	11.8	7.88	26.1	21.7	26.3	39.9	27.5	40.2	15.06	21.29	-37.12	-61.28
	0.2	12.90	7.80	10.88	7.26	9.25	4.35	63.2	60.9	63.7	61.4	64.2	61.9	33.50	31.63	-99.57	-98.30
H ₂ PBO	0.1	12.89	8.99	10.51	8.15	10.02	7.60	48.9	22.8	49.3	23.0	51.9	23.2	26.35	12.77	-75.58	-33.89
	0.15	12.98	9.02	11.6	8.55	11.57	7.36	22.0	27.7	22.2	27.9	22.3	28.2	12.94	15.22	-30.46	-42.00
	0.2	12.49	7.52	10.46	7.41	9.80	6.48	45.7	16.8	46.0	16.9	46.4	17.1	24.69	9.53	-70.41	-24.44
H ₂ ABO	0.1	14.00	8.48	13.54	8.04	10.98	6.03	51.2	42.4	27.7	42.7	51.9	43.0	27.67	22.46	-78.75	-66.85
	0.15	13.65	9.05	10.81	8.33	9.93	5.30	64.2	66.4	64.7	66.9	65.2	67.4	34.14	34.73	-100.9	-106.2
	0.2	13.36	8.64	9.60	7.41	9.41	6.64	68.5	34.2	69.1	34.4	69.6	34.7	36.25	18.36	-108.3	-53.04
H ₂ PPY	0.1	13.04	9.00	12.33	8.47	9.93	6.65	53.1	40.4	53.5	40.7	53.9	41.0	28.51	21.54	-82.54	-63.23
	0.15	14.25	8.65	11.47	7.77	8.85	4.22	94.8	78.6	95.6	79.3	96.3	79.9	49.54	40.62	-152.0	-127.6
	0.2	11.85	8.75	9.92	7.55	8.84	6.19	51.8	44.3	52.2	44.7	52.6	45.0	27.63	23.48	-80.94	-69.99

Table 5. The stability constants of Cu(II) complexes in 50% (v/v) dioxane-water and KCl at different temperatures.

Compound	μ (mol L ⁻¹)	Dissociation constant					
		298 K		303 K		308 K	
		log K1	log K2	log K1	log K2	log K1	log K2
Cu(II) + H ₂ PPS	0.1	18.58	11.30	16.83	9.77	11.36	3.32
	0.15	–	7.36	–	14.16	–	9.93
	0.2	16.20	10.98	14.87	8.59	–	6.57
Cu(II) + H ₂ PBO	0.1	17.71	6.91	15.42	6.38	14.01	3.96
	0.15	18.72	9.85	–	12.43	–	13.21
	0.2	–	13.31	–	12.07	–	13.80
Cu(II) + H ₂ APO	0.1	18.42	9.75	16.06	8.92	14.91	6.43
	0.15	12.52	7.65	–	12.29	9.16	6.96
	0.2	15.93	10.31	–	11.16	–	10.74
Cu(II) + H ₂ PPY	0.1	19.70	11.20	16.29	8.34	15.43	8.23
	0.15	14.68	–	13.37	–	–	7.47
	0.2	14.45	10.64	13.98	11.38	–	6.49

Figure 1. Effect of free ligands and their Cu(II) complexes on the DNA *in vitro*.

data suggest that conjugation between the thio and imine make them inefficient, possibly by reducing the triplet state ($n-\pi^*$) and/or the $^3(\pi-\pi^*)$ lifetime of the sulfur ligand in an enolic form. The absence of such conjugation in **6**, **7**, **8**, and **9** in addition to the near planarity of the chelate ring enhances the photosensitization and consequently can either cause damage of DNA directly or activate oxygen. Therefore, **7** and **8** can be used as a promising anti-tumor agent *in vivo* to inhibit the DNA replication in the cancer cells and not allow the tumor to further grow [67].

4. Conclusion

Based on the elemental analysis and spectral data for Cu(II) complexes of H₂PPS, H₂PBO, H₂APO, and H₂PPY, the substituents modify the coordination. The ligands

coordinate to copper as SN bidentate, NON tridentate, or NSNS tetradentate, forming structures in which each copper is square-planar or octahedral. ESR spectra of these complexes are quite similar, exhibiting axially symmetric g -tensor parameters with $g_{\parallel} > g_{\perp} > 2.0023$ revealing $d(x^2-y^2)$ as ground-state. Thermal degradation indicated high stability of the formed chelates. Also, pH-metric studies were carried out at different temperatures and ionic strengths, and revealed the effect of the substituent on the dissociation constants of the thiosemicarbazides. At a given temperature, pK values are in the order $H_2APO > H_2PPS > H_2PPY > H_2PBO$. DNA degradation assay for H_2PPS , H_2PBO , H_2APO , and H_2PPY and their corresponding Cu(II) complexes showed that the existence of conjugation in the thiosemicarbazides reduced the triplet state lifetime of the sulfur ligand in the thiol form, so they were unable to cause damage on DNA. Absence of such conjugation in the Cu(II) complexes enhanced their ability to cause DNA cleavage. $[Cu(HPBO)Cl(H_2O)](H_2O)$ and $[Cu(HAPO)Cl](H_2O)_3$ have the highest degradation ability.

References

- [1] D.X. West, S.B. Padhyé, P.B. Sonawane. *Struct. Bond.*, **76**, 1 (1991).
- [2] D.X. West, A.E. Liberta, S.B. Padhyé, R.C. Chikate, P.B. Sonawane, A.S. Kumbhar, R.G. Yerande. *Coord. Chem. Rev.*, **123**, 49 (1993).
- [3] M.B. Ferrari, G.G. Fava, E. Lepoati, G. Pelosi, R. Rossi, P. Tarasconi, R. Albertini, A. Boati, P. Lunghi, S. Pinelli. *J. Inorg. Biochem.*, **70**, 145 (1998).
- [4] P.B. Sonawane, R.C. Chikate, A.S. Kumbhar, S.B. Padhyé, R.J. Doedens. *Polyhedron*, **13**, 195 (1994).
- [5] A.M. Thomas, A.D. Naik, M. Nethaji, A.R. Chakravarty. *Inorg. Chim. Acta*, **357**, 2315 (2004).
- [6] M. Belicchi-Ferrari, F. Bisceglie, G. Pelosi, P. Tarasconi. *Polyhedron*, **27**, 1361 (2008).
- [7] P.E. Rapheal, E. Manoj, M.R. Prathapachandra Kurup. *Polyhedron*, **26**, 818 (2007).
- [8] H. Beraldo, D. Gambino. *Min. Rev. Med. Chem.*, **4**, 31 (2004).
- [9] M.B. Ferrari, F. Bisceglie, G.G. Fava, G. Pelosi, P. Tarasconi, R. Albertini, S. Pinelli. *J. Inorg. Biochem.*, **89**, 36 (2002).
- [10] H.S. Seleem, B.A. El-Shetary, S.M.E. Khalil, M. Mostafa, M. Shebl. *J. Coord. Chem.*, **58**, 479 (2005).
- [11] J.R.J. Sorenson. *Prog. Med. Chem.*, **26**, 437 (1989).
- [12] M. Kato, Y. Muto. *Coord. Chem. Rev.*, **92**, 45 (1988).
- [13] J.E. Weder, C.T. Dillon, T.W. Hambley, B.J. Kennedy, P.A. Lay, J.R. Biffin, H.L. Regtop, N.M. Davies. *Coord. Chem. Rev.*, **232**, 95 (2002).
- [14] G. Psomas, A. Tarushi, E.K. Efthimiadou, Y. Aanakis, C.P. Raptopoulou, N. Katsaros. *J. Inorg. Biochem.*, **100**, 1764 (2006).
- [15] D.S. Sigman, D.R. Graham, V.D. Aurora, A.M. Stern. *J. Biol. Chem.*, **254**, 12269 (1979).
- [16] F.Q. Liu, Q. Wang, K. Jiao, F. Jian, G. Liu, R. Li. *Inorg. Chim. Acta*, **359**, 1524 (2006).
- [17] M.M. Meijler, O. Zelenko, D.S. Sigman. *J. Am. Chem. Soc.*, **119**, 1135 (1997).
- [18] S.A. Ross, M. Pitié, B. Meunier. *Eur. J. Inorg. Chem.*, **1999**, 557 (1999).
- [19] G. Psomas, C.P. Raptopoulou, L. Iordanidis, C. Dendrinou-Samara, V. Tangoulis, D.P. Kessissoglou. *Inorg. Chem.*, **39**, 3042 (2000).
- [20] C. Dendrinou-Samara, G. Psomas, C.P. Raptopoulou, D.P. Kessissoglou. *J. Inorg. Biochem.*, **83**, 7 (2001).
- [21] B.K. Santra, P.A.N. Reddy, G. Neelakanta, S. Mahadevan, M. Nethaji, A.R. Chakravartym. *J. Inorg. Biochem.*, **89**, 91 (2002).
- [22] L.Z. Li, C. Zhao, T. Xu, H. Ji, Y. Yu, G. Guo, H. Chao. *J. Inorg. Biochem.*, **99**, 1076 (2005).
- [23] K.K. Du, S.X. Liu. *J. Mol. Struct.*, **874**, 138 (2008).
- [24] L.G. Van Uitert, C. Hass. *J. Am. Chem. Soc.*, **75**, 365 (1953).
- [25] J. Sambrook, E.F. Fritsh, T. Maniatis. *Molecular Cloning: A Laboratory Manual*, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY (1989).
- [26] N.L. Allinger. *J. Am. Chem. Soc.*, **99**, 8127 (1977).
- [27] *HyperChem (Version 8.0.3)*. Available online at: www.hyper.com (2007).
- [28] T.A. Yousef, F.A. Badria, S.E. Ghazy, O.A. El-Gammal, G.M. Abu El-Reash. *Inter. J. Med. Medical Sci.*, **3**, 37 (2001).

- [29] D.X. West, G. Ertem, M. Patricia, J.P. Scovill, D.L. Klayman, L. Judith, F. Anderson, R. Gillardi, C. George, L.K. Pannel. *Trans. Met. Chem.*, **10**, 264 (1985).
- [30] Z. Li, Y. Zhang, Y. Wang. *Phosphorous, Sulphur, Silicon Relat. Elem.*, **178**, 293 (2003).
- [31] A. Braibanti, F. Delavalla, M.E. Pellinghelli, E. Laporti. *Inorg. Chem.*, **7**, 430 (1968).
- [32] I.M. Bazova, R.G. Dubenkon, P.S. Pel'kis. *J. Organ. Khimii*, **14**, 195 (1976); *Chem. Abstr.*, **84**, 105468w (1976).
- [33] D.K. Rastoh, K.C. Sharma. *J. Inorg. Nucl. Chem.*, **36**, 2219 (1974).
- [34] M.J.M. Campbell. *Coord. Chem. Rev.*, **15**, 279 (1975).
- [35] R. Mayer, M. Jansen (Eds). *Organosulfur Chemistry*, Interscience, New York (1967).
- [36] M.S. Refat. *J. Mol. Struct.*, **842**, 24 (2007).
- [37] R. Carpignano, P. Savarino, E. Barni, G. Viscardi. *J. Heterocycl. Chem.*, **21**, 561 (1984).
- [38] F.A. El-Saied, A.A. El-Asmy, W. Kaminstry, D.X. West. *Trans. Met. Chem.*, **28**, 954 (2003).
- [39] S. Delgado, M. Moran, U. Fernandez. *J. Coord. Chem.*, **12**, 105 (1982).
- [40] D. Kivelson, R. Niemen. *J. Chem. Phys.*, **35**, 149 (1961).
- [41] G. Speie, J. Csihony, A.M. Whalen, C.G. Pie-pont. *Inorg. Chem.*, **35**, 3519 (1996).
- [42] B.J. Hathaway, D.E. Billing. *Coord. Chem. Rev.*, **5**, 143 (1970).
- [43] J.L. Mesa, J.L. Pizarro, M.I. Arriortua. *Cryst. Res. Technol.*, **33**, 3 (1998).
- [44] V.T. Kasumo. *Spectrochim. Acta, Part A*, **57**, 1649 (2001).
- [45] (a) H. Yokoi, M. Chikira. *J. Am. Chem. Soc.*, **97**, 3975 (1975); (b) V.H. Growford, W.E. Hatfield. *Inorg. Chem.*, **16**, 1336 (1977).
- [46] S. Pal. *Proc. Ind. Acad. Sci. (Chem. Sci.)*, **114**, 417 (2002).
- [47] O. Khan, T. Mallah, J. Gouteron, S. Jeanin, Y.J. Eanin. *J. Chem. Soc., Dalton Trans.*, 1117 (1989).
- [48] A.W. Addison. In *Spectroscopic and Redox Trends from Model Copper Complexes*, K.D. Karlin, J. Zubietta (Eds), p. 460, Adenine Press, New York (1983).
- [49] V.V. Pavlischuk. *Theor. Exp. Chem.*, **31**, 1 (1995).
- [50] R.K. Ray, G.R. Kauffman. *Inorg. Chim. Acta*, **173**, 207 (1990).
- [51] K. Jayasubramanian, S.A. Samath, S. Thambidurai, R. Murugesan, S.K. Ramalingam. *Trans. Met. Chem.*, **20**, 76 (1995).
- [52] V.S.X. Anthonisamy, R. Murugesan. *Chem. Phys. Lett.*, **287**, 353 (1998).
- [53] V.S.X. Anthonisamy, R. Anantharam, R. Murugesan. *Spectrochim. Acta, Part A*, **55**, 135 (1999).
- [54] R.C. Chikate, A.R. Belapure, S.B. Padhye, D.X. West. *Polyhedron*, **24**, 889 (2005).
- [55] N. Raman, Y.P. Raja, A. Kulandaisamy. *Proc. Ind. Acad. Sci. (Chem. Sci.)*, **113**, 183 (2001).
- [56] R.P. John. *Spectrochim. Acta, Part A*, **59**, 1349 (2003).
- [57] G.A.A. Al-Hazmi, M.S. El-Shahawi, E.M. Gabr, A.A. El-Asmy. *J. Coord. Chem.*, **58**, 713 (2005).
- [58] N.M. El-Metwally, E.M. Gabr, A.A. Abou-Hussen, A.A. El-Asmy. *Trans. Met. Chem.*, **31**, 71 (2006).
- [59] O.A. El-Gammal. *Spectrochim. Acta, Part A*, **75**, 533 (2010).
- [60] S. Sagdinc, B. Koksoy, F. Kandemirli, S.H. Bayari. *J. Mol. Struct.*, **917**, 63 (2009).
- [61] H.M. Irving, H.S. Rossotti. *J. Chem. Soc.*, 2904 (1954).
- [62] M. Jelkic, D. Veselinovic, P. Djurdevic. *Talanta*, **59**, 655 (1992).
- [63] M.G. Abd El-Wahed, S.M. Metwally, M.M. El-Gamel, S.M. Abd El-Haleem. *Bull. Kor. Chem. Soc.*, **22**, 663 (2001).
- [64] A.M. Thomas, A.D. Naik, M. Nethaji, A.R. Chakravarty. *Inorg. Chim. Acta*, **357**, 2314 (2004).
- [65] M. Baldini, M.B. Ferrari, F. Bisceglie, G. Pelosi, S. Pinelli, P. Tarasconi. *J. Inorg. Chem.*, **42**, 2049 (2003).
- [66] Q. Xuhong, H. Tian-Bao, W. Dong-Zhi, Z. Dong-Hui, F. Mingcai, V. Wei. *J. Chem. Soc., Perkin Trans.*, **2**, 715 (2000).
- [67] U. El-Ayaan, M.M. Youssef, S. Al-Shihry. *J. Mol. Struct.*, **936**, 213 (2009).